

## Bio- Synthesis Of Silver Nanoparticles From Leaf Extracts Of *Martynia Annu L.* And Its Antimicrobial Activity.

S.R.Sivakumar\* , C.Sasikala<sup>1</sup> And T.Sheela<sup>2</sup>

\*Assistant Professor, Department of Botany, Bharathidasan University, Trichy-24, SouthTamilnadu, India.

<sup>1</sup>Department of Biotechnology, Dhanalakshmi Srinivasan College of Arts and Science for Women, Perambalur - 621212, NorthTamilnadu, India.

<sup>2</sup>Assistant Professor, Department of Biotechnology, Dhanalakshmi Srinivasan College of Arts and Science for Women, Perambalur -621212, NorthTamilnadu, India.

\* Corresponding Author : S.R.Sivakumar

---

**ABSTRACT:** In the present study Silver nanoparticles were synthesized from aqueous leaf extract of *Martynia annua L.* and characterized by UV-Vis spectroscopy, Fourier transform infrared spectroscopy (FTIR), X-Ray Diffraction (XRD), Scanning electron microscopy (SEM) and Energy Dispersive X(EDX). Characterization by the above said instrument analysis confirmed the presence, size and stability of silver nanoparticles. After characterization, the silver nanoparticles were tested at 100 $\mu\text{g}^{-\text{ml}}$ , 200 $\mu\text{g}^{-\text{ml}}$ , 300 $\mu\text{g}^{-\text{ml}}$  and 400 $\mu\text{g}^{-\text{ml}}$  concentrations to check the bactericidal activity against clinical isolates of five bacterial pathogens. We observed that, if the concentration of leaf extracts synthesized nanoparticle increases, the zone of inhibition also get increased in all the test five clinical bacterial pathogens against streptomycin and result suggested the potential use of leaf extracts synthesized silver nanoparticles against other clinical pathogens.

**KEY WORDS:** *Martynia annua* leaf extracts, silver nanoparticle synthesis, characterization, antibacterial activity against clinical pathogens.

---

Date of Submission: 19-05-2018

Date of acceptance: 04-06-2018

---

### I INTRODUCTION

Nanotechnology can be delineated as a research for the design, synthesis, and manipulation of structure of particles with dimension smaller than 10 nm. The idea of nanotechnology was first introduced by (Richard Feynman in 1959). The popular term "Nanotechnology" was coined by Norio Taniguchi, a Japanese professor of Tokyo Science University, in 1974 engineer materials at nanometer level. Different types of Nano-Materials are present in Quantum Dots, Nanocrystals, Nanoparticles, Metallic Nanoparticles, Dendrimers, Bucky balls, and Nanotubes. Nanotechnology is an interdisciplinary field and is formed from the convergence of chemistry physics and biology.

Nanoparticles show completely new or improved properties such as size distribution and morphology of the particles etc. Novel application of nanoparticles and nonmaterial are emerging rapidly field (Kaviya et al., 2011). As motioned above, generally silver is used in the form of nitrate to induce antimicrobial effect. But when silver nanoparticles are used there is huge increase in surface area available for the microbe to be exposed to. Though silver nanoparticles find use in many antibacterial applications the action of this metal on microbes is not fully known. Silver nanoparticles have long been known for their antibacterial, antifungal, antiviral and anti-inflammatory properties. In medicines silver is used to prevent infection of burns and open wounds (Duran et al., 2005). Medical devices and implants are prepared with silver impregnated polymers (Ro et al., 1999). In textile industry silver embedded fabrics are now used in sporting equipment (Klaus et al 1999). Biological methods are ecofriendly and are cost effective.

The pure metals in nanoparticles form are applied in the field of diagnostics antimicrobial agents, drug delivery, textiles (clothing) electronics, bio sensing food industry paints, cosmetics, medical devices and treatment of several acute and chronic diseases like – malaria, hepatitis, cancer and aids due to various events occurring in the environment among them human activities of prime importance that result production of reactive oxygen species (ROS) showing negative impact in biological system. The progress in time evidenced development in technology that revealed the ability metals of nanoscale to perform specific functions better than the bulk form of metals. The noble metals like silver gold, platinum palladium copper, zinc and iron were used in synthesis of particles of nano size. The nanoparticles such as size shape, composition, crystalline nature and structure determine their applications. Nanoparticles involve algae, actinomycetes, bacteria, fungi and plants.

*Martynia annua L.* belongs to the family Martyniaceae. The plant is commonly known as Devil's claw (English) Bichu (Hindi) Kakanasika (Sanskrit) Vechhida (Gujarati) and tamil name buli nagam. It is a small herbaceous, erect, branched, glandular hairy annual herb growing up to 0.9- 1.2 m in height leaves are large simple opposite green in color broadly ovate to triangular ovate, glandular hairy 9-22 x 9-20 cm cordate at base branched and covered with glandular hairs. Flowers are drooping large pale mauve or lavender in short spikes at the end of branches. They are tubular shaped 4-6 cm long pink and dark purple blotched with yellow inside foxglove shaped ill-smelling and terminate in 5 spreading lobes with a prominent spot between each. The plant is used for the treatment in Ayurveda. The plant is known as kakanasika. This is being used in Indian traditional medicines for epilepsy, inflammation and tuberculosis. The leaves of *Martynia annua* are edible and used antiepileptic and antiseptics applied locally to tuberculosis glands of the neck the juice of the leaves as a gargle for sore throat and the leaf paste for wounds of domestic animal. The leaf is used to kill bugs. The leaf paste is antidote to venomous strings and applied to tuberculosis glands. The leaf juice is used in gargle treatment. The methanolic extract of *Martynia annua* leaves shows the presence of higher amount of terpenoid alkaloids glycosides steroids tannins, saponins and moderate amount of p-hydroxyl benzoic acid, synaptic acid and fatty acid such as plasmatic acid and stearic acid.

The present study explores the pharmacological potential of the plant. The chloroform, ethyl acetate and methanol extract of *Martynia annua* leaves were found to have antibacterial activity. When evaluated against gram positive and gram negative bacteria. All the solvent extracts show antibacterial action respective to different bacteria. Chloroform extract shows higher antibacterial activity against *Proteus vulgaris*, *Bacillus subtilis* and *B. thuringensis*. Ethyl acetate extract was potentially effective against *Salmonella paratyphoid*, *A. Salmonella paratyphoid B*, *Proteus mirabilis* the methanol fraction of ethanol extract of *Martynia annua* leaves shows significant wound healing effect by stimulating of wound contraction as well as epithelialization.

Moreover, phytochemical studies demonstrated that. The methanol fraction mainly contains flavonoid luteolin responsible for enhancement of wound healing process due to the free radical scavenging mechanism. It is imperative that more clinical and pharmacological trials are needed to investigate the unexploited potential of this plant.

In the present study synthesis of silver nanoparticles using the leaves extract of the plant *Martynia annua*. To characterize the silver nanoparticles by UV-VIS, FTIR, SEM, EDAX and XRD and to find out the antimicrobial activity of the synthesized silver nanoparticles against clinical pathogens.

## II MATERIALS AND METHODS:

### MATERIALS AND METHODS:

#### PLANT COLLECTION:

Fresh leaves *Martynia Annu L.* were collected at Bharathidasan University campus at Trichy, Tamil Nadu India waste places and road sides. Plant was authenticated from (Dr.S.Joh brittle) St. Joseph College (Campus) Tiruchirappali.

#### PREPARATION OF LEAF EXTRACT

The fresh leaves were washed with running tap water for 20 mins. and dry in shade at room temperature for one week. Then the leaves are cut into small pieces and make into fine powder. 20g of leaf extract are weighed and dissolved in 200ml distilled water in a 500ml Erlenmeyer flask and boil for 30mins. The extract was filtered with Whatman No.1 filter paper and stored in an airtight container under dark condition until for further use.

#### PREPARATION OF SILVER NANOPARTICLES

1mM of silver nitrate ( $\text{AgNO}_3$ ) was prepared in 1000ml beaker (0.1698 g  $\text{AgNO}_3$  is added to 1000ml of distilled water). The 100ml leaf extract were mixed with 900ml silver nitrate solution (1:9) ratio. and kept under dark condition. Colour change of the solution from white with pale yellow indicated that the silver nanoparticles get synthesized and then the solution was centrifuged at 7,000rpm, 20°C for 15mins. Then the supernatant was collected from the tube and it was kept for evaporation (to sediment the particles) until it gets fully evaporated. Collect the pellet which is kept in hot air oven at 40°C for twenty four hours. Further, the synthesized sample was used for characterization and antimicrobial activity studies.

#### CHARACTERIZATION OF SILVER NANOPARTICLES

The characterization of silver nanoparticles was carried out by different instrument and technique. It includes visual observation, UV- Vis Spectrophotometer, FTIR, XRD, SEM, and EDAX.

### **UV- VIS Spectrophotometer**

To determine the time point of maximum production of silver nanoparticles, the absorption spectra of the samples were taken 300 to 600 nm using a UV–Vis spectrophotometer. The deionised water was used as the blank.

### **FOURIER TRANSFORM INFRAED SPECTROSCOPY (FTIR)**

FTIR analysis were carried out to identify the possible bio molecules responsible for reduction of the  $\text{Ag}^+$  ions and capping of the bio reduced silver nanoparticles synthesized.

### **X-Ray diffraction analysis (XRD)**

The X-ray diffraction pattern indicated the crystalline structure of silver nanoparticles. The XRD spectrum confirmed the presence of silver nanoparticles. The diffracted intensities were recorded from  $2\theta$  angle.

### **SEM (Scanning Electron Microscope) analysis**

The silver nanoparticles were also characterized by scanning electron microscopy (SEM). The direct electron microscopic visualization allows measuring the size and shape of biocapped silver nanoparticles formed.

### **EDAX (Energy- Dispersive X-ray Spectroscopy) analysis**

EDAX is an analytical technique used for the elemental analysis or chemical characterization of a sample. It relies on an interaction of some source of X-ray excitation and a sample. Its characterization capabilities are due in large part to the fundamental principle that each element has a unique atomic structure allowing unique set of peaks on its X –ray spectrum. To stimulate the emission of characteristic X-rays from a specimen, a high-energy beam of charged particles such as electrons or protons (see PIXE), or a beam of x ray, is focused into the sample being studied. The number and energy of the X rays emitted from a specimen can be measured by an energy –dispersive spectrometer.

### **Antibacterial activity studies**

Silver nanoparticles synthesized from *Martynia Annu* leaves were screened for antibacterial activity against clinical pathogenic bacteria namely *Escherichia coli*, *Pseudomonas aeruginosa*, *Rhodococcus rhodnii*, *Streptococcus aureus* and *Proteus mirabilis*

### **Media preparation:**

Nutrient broth (Peptone – 5g, Beef extract – 1.5g, Yeast extract – 1.5g, NaCl – 5g and Distilled water – 600ml; pH adjusted to 7.2) was prepared. After sterilization of the medium, the bacterial culture was inoculated in the nutrient broth. The inoculated broth has been incubated for 24 hours at 37°C in incubator. Nutrient agar (Peptone – 5g/l, Yeast extract – 1.5g/l and Beef extract – 1.5g/l, pH of 7.2) was prepared, sterilized and poured on to the sterilized petriplates.

### **Preparation of inoculums:**

Bacterial inoculums were prepared by transferring a loop full of bacterial culture from fresh culture plates to tubes containing 10 mL of Nutrient Broth (Hi-media) and incubated for 24 hours at 37°C. The tubes were shaken occasionally to aerate and promote growth. These cell suspensions were diluted with sterile Nutrient Broth to provide initial concentration cell counts of about  $2 \times 10^3$  CFU<sup>-mL</sup> at 600nm O.D at Spectrometer. After the solidification of the media in the petriplate, bacterial cultures were inoculated by swap method.

### **Disc diffusion method**

The antibacterial activity of crystalline bio molecule capped synthesised silver nanoparticles from *Lobophora variegata* was determined by disc diffusion method. Discs of 6mm diameters were prepared from Whatmann No.1 Filter paper and kept in the hot air oven at 160°C for 1 hour. The nutrient agar plates were prepared and inoculated with test bacterial organisms by spreading the bacterial inoculums on the surface of the media. The discs were impregnated with different concentrations ranging of 100 $\mu\text{g}^{-\text{ml}}$ , 200 $\mu\text{g}^{-\text{ml}}$ , 300 $\mu\text{g}^{-\text{ml}}$  and 400 $\mu\text{g}^{-\text{ml}}$ . A negative control was prepared by taking 1mM Silver nitrate dissolved in 1ml distilled water and positive control Streptomycin used as positive controls (100  $\mu\text{g}^{-\text{ml}}$ ).

### **Evaluation of antibacterial activity**

The plates were incubated at 37°C for 24 hours. The antibacterial activity was assessed by measuring the diameter of the area in which bacterial growth was inhibited around the disc and measured the diameter zone of inhibition (in mm).

### **III RESULT AND DISCUSSION:**

Synthesis of silver nanoparticles:

Biological and biomimetic approaches for the synthesis of nanomaterials are being explored. Cell mass or extracellular components from microorganisms, such as *Klebsiella pneumonia*, *Bacillus licheniformis*, *Fusarium oxysporum*, *Aspergillus flavus*, *Cladosporium cladosporioides*, *Aspergillus clavatus*, and *Penicillium brevicompactum*. The unexploited plant resources for the synthesis of silver nanoparticles, various plant leaf extracts such as *Helianthus annuus*, *Basella alba*, *Oryza sativa*, *Saccharum officinarum*, *Sorghum bicolor* and *Zea mays*, *Jatropha curcas* and *Aloe vera*.

Development of reliable and eco- friendly process for synthesis of metallic nanoparticles is an important step in the field of application of nanotechnology. The principle of preparation of silver nanoparticles by using microorganism is a bioreduction process the silver ions are reduced by the extracellular reductase enzymes produced by the microorganisms to silver metal in nanometer range. Synthesis of advanced materials energy storage devices, electronic and optical displays chemical and biosensors as well as biomedical devices, recognizing the important of nanomaterials in key future technology.

Synthesis of nanoparticles using microorganisms involves elaborate process of maintaining cell cultures, intracellular synthesis and multiple purification steps. In this regard using “green” methods in the synthesis of zinc oxide nanoparticles has increasingly become a topic of interests as conventional chemical methods are expensive and re-quire the use of chemical compounds/organic solvents as reducing agents (Cynthia Mason et al., 2012). Plants have a lot of phytochemicals in their parts: they are applied in various fields. The biochemicals may play an important role in the nanoparticles synthesis (T.Santhoshkumar, et al.,2011) *Solanum trilobatum* Linn is an important medicinal plant of the family solanaceae. The leaves contain rich amount of calcium, iron, phosphorus, carbohydrates fat, crude fiber, and minerals (M.Jawhar, et al.,2004). This plant has strong immune stimulatory effect due to the presence of alkaloids and carbohydrates. Due to these antioxidant and antibiotic properties this was used to synthesize silver nanoparticles.

In this present investigation the medically important plant is used for the synthesis of medically valued silver nanoparticles. The morphological, crystalline, and biochemical characters of green synthesized silver nanoparticles were analyzed by UV-VIS spectrophotometer, Scanning Electron Microscope, X-ray Diffraction pattern, and Fourier Transform Infrared spectroscopy. Finally the medical property of the silver nanoparticle was characterized using antibacterial assay. Many plants are used for synthesizing nanoparticles including *Cinnamomum camphora*, (Huang et al., 2007) Preparation and synthesis of silver nanoparticles from the callus and in-vitro derived plant roots of white flowered variety of *Clitoria ternatea*. Few papers reported the biosynthesis of zinc oxide nanoparticles using plant extracts such as *Aloe barbadensis* M.(Sangeetha.,2011). *Parthenium hysterophorus* (Rajiv P et, al 2014) and *Poncirus trifoliata* plant dried fruits (Nagajyothi.,et al.,2013) and milky latex of *Calotropis procera* (Singh R,et al 2011).

*Martynia annua* extract used to produce silver nanoparticles in this experiment Ag<sup>+</sup> ions were reduced to Ag nanoparticles when plant extract was mixed with AgNO<sub>3</sub> solution in 1:8 ration reduction was followed by on immediate change in yellowish to brown color in the aqueous solution of the plant extract due to surface Plasmon vibration in silver nanoparticle.

Plant was mixed with AgNO<sub>3</sub> solution. The mixture was kept at room temperature for 24 hours. The appearance of yellowish brown color in the reaction vessel indicated formation of AgNO<sub>3</sub> AgNO<sub>3</sub> exhibit this yellow-brown color in aqueous solution due to excitation of surface Plasmon resonance in the AgNO<sub>3</sub>. Primary conformation of the AgNO<sub>3</sub> was carried out by UV-Visible spectrophotometric analysis. The nanoparticles showed maximum absorbance peak at 380 nm on UV-Vis spectra which is shown in Fig 1. The UV-Vis spectra recorded from the *Martynia annua* reaction vessel at the strong surface Plasmon resonance centered at 380 nm clearly showed.

### **FTIR spectrum:**

The FTIR spectrum of extracellular biosynthesized AgNO<sub>3</sub> was shown in Fig FTIR measurement of the freeze – dried samples were carried out to identify the possible interactions between silver and bio-molecules which may be responsible for synthesis and stabilizations of silver nano particles with capping agent available in the *Martynia annua* broth.

The amide linkages between amino acid residues in protein give rise to well known signatures in the infrared region of the electromagnetic spectrum. The FTIR spectrum revealed five bands at 3429.8 cm present the O-H stretch H- bonded vibration of alcohols, and Phenols.

#### **Scanning Electron Microscope (SEM) Analysis:**

The SEM analysis was used to determine the structure of the reaction products that were formed Fig6 the silver nanoparticles observed were of spherical shape of the scanning electron microscope the nanoparticles were analyzed in the different magnifiers.

#### **EDAS:**

In the present study, for the conformation of AgNO<sub>3</sub>, EDAX spectroscopy analysis was performed which confirmed the presence of elemental silver by the sharp signals.

#### **X-Ray Diffraction (XRD) Analysis:**

The powdered sample was used by a CuK $\alpha$ -X Ray Diffract meter for confirming the presence of silver nitrate and analyzing the structure. The graph showed main peaks main peaks corresponding to 2 $\theta$  values of 37.5, 46.5 and 77.8 correspond to (111)(200)(311) in the multi-plot shown in Fig 6. The location of the peaks was compared to literature values and the presence of silver nitrate particles was confirmed. Similar results were reported earlier in silver nanoparticles.

#### **Antimicrobial activity:**

In the present study the assay for antimicrobial activity was done against escherchia coli, salmonella typhimurium, Vibrio choleraera Pseudomonas aeruginas and pseudomonas mirabilis with various concentration (100,200,300 and 400 /ml) of green synthesized silver nanoparticles of Martynia annua and the result are showed in fig.7. The silver nanoparticles showed good activity against all tested organisms at all concentration. The highest zone of inhibition was found against *Vibrio choleare* (19mm at 400) and minimum level of antibacterial activity was observed against *pseudomonas aeruginosa* (10mm at 100). The above observation clearly indicated that the synthesized silver nanoparticles have the potential to kill the bacteria effectively.

### **IV SUMMARY AND CONCLUSION**

The biological synthesis of silver nanoparticles using Martynia annua leaves extract provides environmental friendly and efficient method for synthesis of nanoparticles. The synthesized nanoparticles were of spherical shape and the estimated size. The size was bigger as the nanoparticles were surrounded by a thin layer of metabolites. Which were found from the characterization using UV-vis spectrophotometer, FTIR, EDAS, SEM and XRD techniques. All the above analyses proved that the concentration of plant extract to metal iron ration plays in important role into the shape determination of the nanoparticles.

The synthesized silver nanoparticles exhibited good anti bacterial activity against clinical isolates of five bacterial pathogens. The present study offers the ability of *Martynia annua L.* leaf extract to reduce silver ions to synthesis silver nanoparticles which can be used for several applications like anti-bacterial, antifungal activities and further LC-MS characterization is to be done for organic compound capped in it. From the technological point of view these silver nanoparticles have potential application in the biomedical field and this simple procedure has several advantages such as cost- effectiveness, compatibility for medical and pharmaceutical applications as well as large scale commercial production.



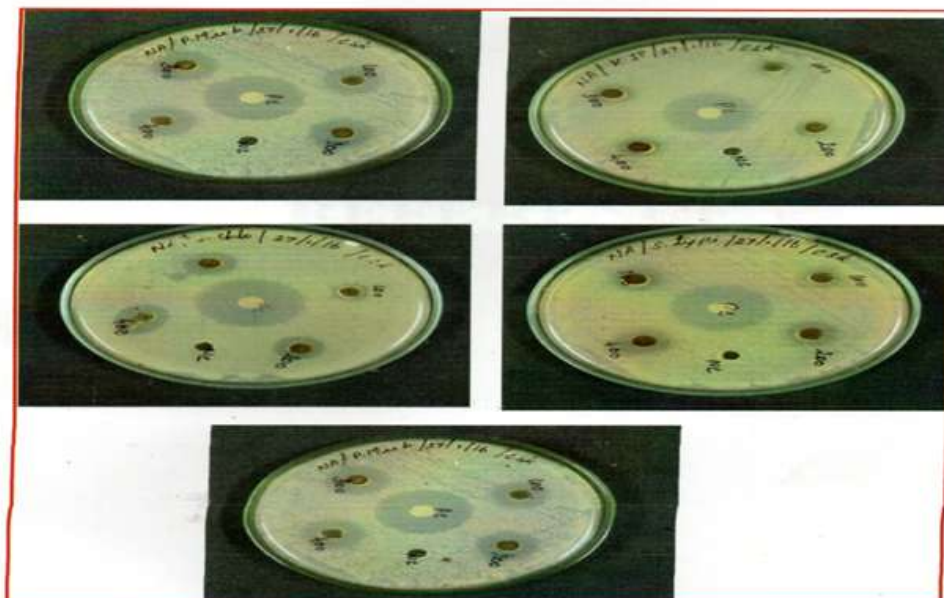
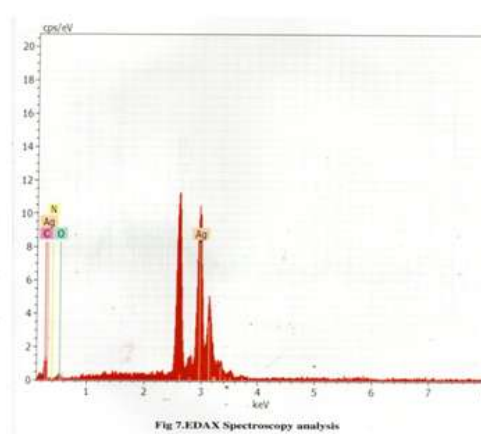
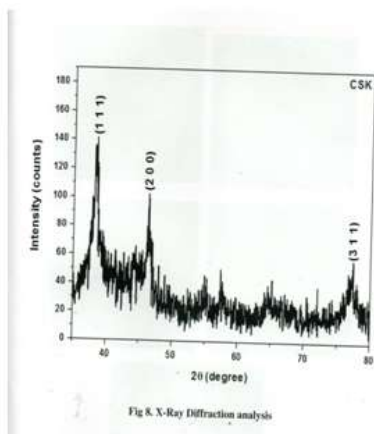
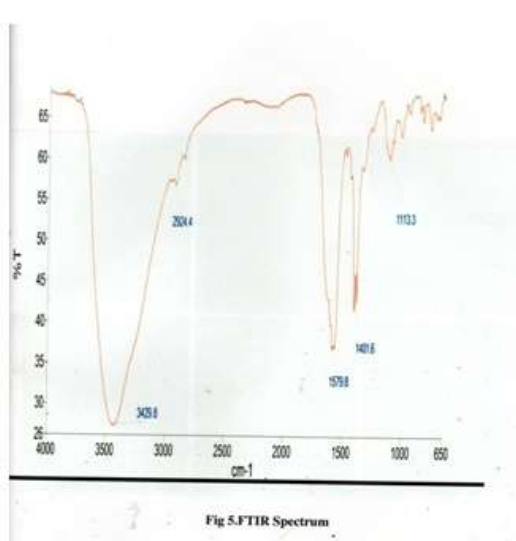
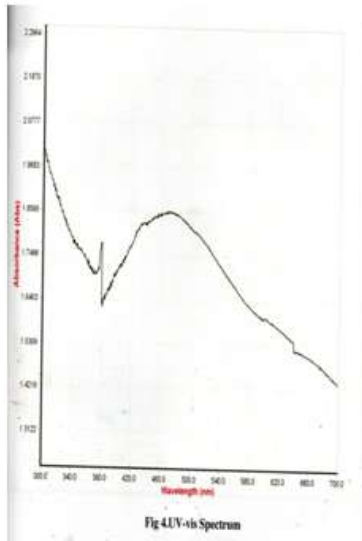


Fig 9. Antimicrobial activity of *Martynia annua* silver nanoparticles



**Table 1: Antimicrobial activity of *Martynia annua* silver Nanoparticles**

Sample concentration (µg/ml)	Diameter of Zone inhibition (mm)				
	<i>E.coli</i>	<i>Salmonella typhymurium</i>	<i>Vibrio cholerae</i>	<i>Psudomonas aeruginosa</i>	<i>Psudomonas mirabilis</i>
100	14	11	12	10	14
200	16	13	14	13	15
300	17	15	18	15	16
400	18	18	19	17	18

**Table 2: FTIR Spectroscopy analysis**

FREQUENCY cm	BOND	FUNCTIONAL GROUP
3429.8	O-H stretch – bond	Alcohols phenols
2924.4	C-H stretch	Alkanes
1579.8	C-C stretch (in-ring)	Aromatics
1401.6	C-H bend	Alkanes
1113.3	C-N stretch	Aliphatic amines

**REFERENCE:**

- [1]. Abu Bakar Ismail NHH and Abu Bakar M, Synthesis and characterization of silver nanoparticles in natural rubber. Mater chen phys, 104:276-283, (2004)
- [2]. Ahmad A, Senapati S, Khan MI, Kumar R and Sastry M, Extracellular biosynthesis of monodisperse gold nanoparticles by a novel extremophilic actinomycete, Thermomonospora sp. Langumir, 19:3350-3553(2003)
- [3]. Bar H, Bhui DK, Sahoo GP, Sarkar P, De SP and Misra A, Green synthesis of silver nanoparticles using latex of jatropha curcas. Colloids and surface A: Physicochemical and Engineering Aspects 339:134-139 (2009)
- [4]. Begum NA, M SaBLRA (2009) Mondal Colloids and surface B. Biointerface 71:P 13-118.
- [5]. Begum NA, Modal S, Baus S Laskar RA and Mandal D, Colloids and Surfaces B: Biointerfaces 113-118, (2009, B)
- [6]. Bera RK, Dad AK Raj CR (2010) Scope of network polysilanes in the synthesis of fluorescent silver and gold nanoparticles/nanoclusters- modulations of their optical properties in the presence of Hg(II) ions. Chem. Mater 22:4505-4511
- [7]. Bhainsa KC And D Souza SF, Extracellular biosynthesis of silver nanoparticles using the fungus Aspergillus fumigates colloids and surfaces B: Biointerfaces 47:160-164, (2006).
- [8]. Bhattacharya D and Ranjinder G, Nanotechnology and potential of microorganisms. Crit Rev Biotechnol, 25:199-204, (2005)
- [9]. Chandran SP, Chaudhury M, Pasricha R, Ahmad A, Sastry M (2006) Synthesis of gold nanoparticles and silver nanoparticles using Aloe vera plant extract. Biotechnology Progress 22(2):577-583.
- [10]. Chandrasekharan N and Kamat PV, Improving the photo electrochemical performance of nanostructured TiO<sub>2</sub> films by adsorption of gold nanoparticles J Phys Chem B, 104:10851-10857, (2000).
- [11]. Cheng Y, Yin Lin S, Wiesner M, Bernhardt E, Liu J (2011) Toxicity reduction of polymer stabilized silver nanoparticles by sunlight J. Phys Chem C, 115:4425-4432.
- [12]. Colvin, VLS, MC & Alivisatos, A (1994) Light emitting diodes made from cadmium selenide nanocrystals and a semiconducting polymer Nature 370: P354-357.
- [13]. Cynthia Mason, Singaravelu Vivekanandhan, Manjusri Misra, Amar Kumar Mohanty (2012) Switchgrass (*Panicum Virgatum*) Extract Mediated Green Synthesis of Silver Nanoparticles World Journal of Nano Science and Engineering Vol 2, pp 47-52
- [14]. Esumi K, Takafumi T, Kanjiro T and Kenjiro M, Preparation and characterization of bimetallic palladium copper colloids by thermal decomposition of their acetate compounds in organic solvents. Chem Mater. 2(5): 564-567 (1990)
- [15]. Farooqui AMD Chauhan PS Moorthy SN Shaik PK (2010) J Extraction of silver nanoparticles from the leaf extract *Clerodendrum icerme* Digest J. Nanomater Biostruct 5:43-49.
- [16]. Fierascu RC, Ion RM and Dumitriu (2008) Plasmonic Materials for Biomedical Application “International Conference 6<sup>th</sup> workshop on European Scientific and Industrial Collaboration on promoting Advanced Technologies in manufacturing WESIC 08, Bucharest, Romania, .
- [17]. Geerthalakshmi R, Sarada DVL (2010) Synthesis of plant mediated silver nanoparticles using *Trianthema decandra* extract and evolution of their anti microbial activities. International Journal of Engineering Science and Technology 2(5) 970-97
- [18]. Gericke M, A Pinches (2006b) Biological synthesis of metal Nanoparticles. Hydrometallurgy 83:132-140
- [19]. Gnanasangeetha D & Sarala TD (2013a) One pot synthesis Zinc oxide nanoparticles via chemical and green method. Research journal of Material science 1(7): 1-8
- [20]. Henglein A, Refuccion fo Ag(CN)<sub>2</sub> on silver and platinum colloidal Nanoparticles Langmuir, 17:2329-2333(2001)
- [21]. Hirsch T, Zhamnikov M, Shaporenko A, Stahl J, Weiss D, Wolfbeis OS et al (2005) size controlled electrochemical synthesis of metal nanoparticles on monomolecular templates, Angew Chem Int. Ed 44:6775-6778.
- [22]. J. Huang, Q. Li, D. su et al., “Biosynthesis of silver and gold nanoparticles by novel sundried cinnamomum camphora leaf” Nanotechnology vol 18, no10, Article ID 105104, 2007.
- [23]. Jha A, Prasad K, Prasad, K, Kulkarni AR (2009) Plant System nature’s nanofactory colloids surf. B. Biointerfaces 73:P.219-223.
- [24]. Kaviya S, SJB Viswanathan B (2011) Green synthesis of silver nanoparticles using *polyalthialongifolia* leaf extract along with D-Sorbitol Journal of Nanotechnology p.1
- [25]. Kartikar KR, Basu BD, Indian medicinal Plants 2<sup>nd</sup> ed vol III, International book distributors deharadun 1987, 1854-1855
- [26]. Klaus T, JR Olsson E, Granqvist CG (1999) Silver based crystalline nanoparticles Microbial Fabricated Pro Nati Acadsc USA 96 P.13611-13614
- [27]. Korotchenkov OA, Cantarero A, Shpak AP, Kunitskii YA Senkavich AI, Borovoy MO et al. (2005) Doped ZnS:Mn nanoparticles obtained by sonochemical synthesis Nanotechnology 16:2033-203
- [28]. Krolkowski A, Kudelski A, Michota A and Bukowska J, SERS studies on the structure of thioglycolic acid monolayers on silver and gold. Surf Sci, 532:227-232(2003)
- [29]. Kumar V and Yadav SK (2009) J. Chem Techn Biotechn, Vol 84, No. 2, P.151.
- [30]. Lim KT, Hwang HS, Ryoo W, Johnston KP (2004) Synthesis of TiO<sub>2</sub> nanoparticles utilizing hydrated reverse micelles in CO<sub>2</sub> Langmuir 20:2466-2472
- [31]. SM, Lin FQ Guo HQ, Zhang ZH and Wang ZG Surface states induced photoluminescence from Mn<sup>2+</sup> doped cds nanoparticles. Solid state common 115:615-618 (2000)
- [32]. Lodhi S, Singhai AK Preliminary Pharmacological evaluation of *Martynia annua* leaves for wound healing. Asian pacific journal of tropical Biomedicine 2011:1(6):421-427

- [33]. M.Jawhar, G.Amalan Rabert, and M.Jeyaseelan, "Rapid proliferation of multiple shoots in solanum trilobatumL, Plant Tissue Culture vol 14,no 2 pp,107-112.
- [34]. Mail PC Anasari AS, Chaturedi M.Antifertilit effect of chronically administered martynia annua root extract on male rats journal of ethnopharmacology 2002;82(2-3) 61-67
- [35]. Masurkar SA, Chaudhari PR, Shidore VB and kamble SP,Rapid Synthesis of silver nanoparticles using C.citrus (Lemongrass) and Its antimicrobial activity. Nano Micro Lett 3:189-194 (2011)
- [36]. Mhaskar KS Blatter E, Caius JF Indian Medicinal plants Vol II Sri Satguru publications delhi, 2000,556-559
- [37]. Nadagouda MN, Speth TF Varma RS (2011) Microwave assisted green synthesis of silver nanostructures Accounts chem. Res 44:469-478
- [38]. Nagda D, Saluja A, Nagda C.Antioxidant Activities of methanolic and aqueous extract form leaves of Martynia annua Linn Jouranal of Pharamacognosy 2009 ;1:288-297
- [39]. Pakinkar KM (2007 Anti microbial activity of Biologically stabilized silver Nanoparticles" pat Appl pub US 0218555A1
- [40]. Parasharu K,SaSa (2009) Bioinspired Synthesis of silver Nanoparticles. Digest Journal of Nanomaterials and Biostructures 4:P 159-166.
- [41]. Pastoriza-Santos and Liz – Marzan LM, Formation of PVP-protected Metal Nanoparticles in DMF Langmuir 18:2888-2894 (2000)
- [42]. Pattabi M,J Uchil (2000) synthesis of cadmium sulphide nanoparticles. Solar energy Mater solar cell 314
- [43]. Plante IJL Zeid TW Yangab P, Mokari T (2010) Synthesis of metal sulfide nanomaterials via thermal decomposition of single – source precursors J.Mater Chem 20:6612-6617
- [44]. Prabhu N,Divya TR, Yamuna G(2010) Synthesis of silver phyto nanoparticles and their antibacterial efficacy. Digest Journal of Nanomaterials Biostructures 5:185-189
- [45]. Rajiv, P.Rajeshwari, S;Venckatesh, R.Spectrochimica Acta part A:Molecular and Bimolecular Spectroscopy 2013, 112, 3847 DOI: 10.1016/j.saa.2013.0407
- [46]. Rashid A, Khaydarov, Renat R Khaydarov Olga Gapurova, Yuri Estrin ThomasvScheper (2009) Electrochemical method for synthesis of silver nanoparticles.J Nanopart Res 11:1193-1200.
- [47]. Rastogi RP Melhotra BN Compendium of Indian Medicinal Plants Vol.II, CDRI, Lucknow, 1993,446/
- [48]. Rodriguez – sanchez L,Blanco MC and Lopez- Quintela MA, Electrochemical Synthesis of Silver Nanoparticles J Phys chem. B,104 (41) 9683-9688(2000)
- [49]. Roh Y,Lauf RJ,Mc Millan AD, Zhang C,Rawn CJ Bai J and Phelps TJ Microbial synthesis and the characterization of metal substituted magnetite solid state commun 1185;529-534,(2001)
- [50]. Rosemary MJ,T Pradeep (2003) Solvothermal synthesis of silver Nanoparticles from thiolates Journal of Colloid and Interface Science 268;81-84
- [51]. Saifuddin N, Wong CW and Yasimura AN, Rapid biosynthesis of silver nanoparticles using culture supernatant of Bacteria with microwave irradiation. E-journal of chemistry 6(1) 61-70 (2009)
- [52]. Sangeetha G, Rajeshwari s & venckatesh R (2011) Green synthesis of Zinc oxide nanoparticles by Aloe barbadensis miller leaf extract; structure and optical properties Materials Research Bulletin 46: 2560-2566
- [53]. Sermakkani M.Thangapandian V.Phytochemical and antibacterial activity of Martynia annua L.against the different pathogenic bacteria . Journal of Herbal Medicine and Toxicology 2010 4(2) : 221-224
- [54]. Sharma PC yelne MB, Dennis TJ (2008) Data base on medicinal plants used in ayurveda Vol, 1, pp 70-71
- [55]. Shekhawat MS & Manokari M (2014) Biogenesis of Zinc oxide Nanoparticles using Morinda pubescens J.E Smith Extracts and their Characterization. International Journal of Bio Engineering and Technology 5 (1) 1-6
- [56]. Shiv Shankar S,Rai A Ahmad A and Sastry M,Rapid synthesis of Ag and Bimetallic Au core – Ag shell nanoparticles using neem (Azadirachta – indica) leaf broth J. Colloid Intef Sci, 275: 496-502 (2004)
- [57]. Singh NP Pnada H Medicinal herbs with their formulation. Daya publishing house new delhi 2005,671
- [58]. Singh R.P Shukla V.K. Yadav R.S Sharma P.K Singh P.K Pandey A.C Adv Mater Lett 2011 2,313 DOI: 10.5185/ amlett india 204.
- [59]. Sivakumar P.Chndran Nethradevi sahadevan Renganthan (2012) Synthesis of silver nanoparticles using Lantana camara fruit extract and its effect on pathogens. Asian journal of Pharmaceutical clinical Research 5.
- [60]. Smetana AB, KJ Klabunde, CM Sorensen (2005) Synthesis of spherical silver Nanoparticles Stabilized with various agents and their 3D and 2D Superlattice Formation J. Colloid Int.Sci 284-521
- [61]. Song JY and kim BS Rapid biological synthesis of silver nanoparticles Using plant leaf extract Bioprocess Biosyst Eng 32(1) 72-84 (2009)
- [62]. Sripriya J, Anandhakumar S, Achirman S, Antony JJ Siva D & Raichur AM (2013) Laser receptive polyelectrolyte thin films doped with biosynthesized silver nanoparticles for antibacterial coating and drug delivery applications. International journal of pharmacy 457 (1) 206-13
- [63]. T.Santhoshkumar A.A. Rahuman G Rajakumar et al "Synthesis of silver nanoparticles using Nelumbo nucifera extract and its larvicidal activity against malaria and filariasis vectors Parasitological Research vol 108 no 3 pp 693-702,2011
- [64]. Taleb C, Petit M and Pileni P, Synthesis of Highly Monodisperse Silver nanoparticles from AOT Reverse Micelles A Way to 2D and 3 D Self organization Chem Mater 9(4) 950-959 (1997)
- [65]. T. Santhoshkumar A.A.Rahuman, G.Rajakumar et al., "Synthesis of silver nanoparticlels using Nelumbo nucifera Extract and its larvicidal activity against malaria and filariasis Vectors Parasitology Research Vol 108, no.3 pp. 693 702, 2011
- [66]. Vigneshwaram N, Nachane RP, Balasubramanya RH and Varadraj PV navel one pot green synthesis of stable silver nano particles soluble starch. Carbohydrate Res 341:2012-2018(2006)
- [67]. Wang Y, HN (1991) Nanometer sized semiconductor clusters material synthesis, quantum sie effects and photophysical roperties J Physchem. 95 p 525-532
- [68]. Watt G. Dictionary of the economic products of india Vol. V cosmo publications New Delhi 1972
- [69]. Willner B, Basnar B and Willner B, Nanoparticle-enzyme hybrid system for nanobiotechnology FEBS J 274 (2) 302-309 (2007)
- [70]. Xu Z P, ZQP, LU G Q and YU A B (2006) Inorganic Nanoparticles As Carriers for Efficient cellular Delivery Chemical Engineering Science 61 P 1027-1040
- [71]. Zhu J J, Liu S W Palchik O, Kolytyn Y and Gedanken A, Shape – Controlled Synthesis of silver Nanoparticles by pulse sono electro chemical Methods Langmuir 16 (16) 6396 – 6399, (2000)

S.R.Sivakumar." Bio- Synthesis Of Silver Nanoparticles From Leaf Extracts Of Martynia Annu L. And Its Antimicrobial Activity " International Journal of Pharmaceutical Science Invention(IJPSI), vol. 07, no. 04, 2018, pp. 40-47.