

## Male reproduction and Cannabis: A state of art review.

Somenath Ghosh\*

\* Corresponding author: Dr. Somenath Ghosh, Assistant Professor, Post Graduate Department of Zoology, Rajendra College, Chapra-841301, Bihar, India.

---

### ABSTRACT

*Cannabis sativa* is a cheap hallucinating agent used in different parts of the world from time unknown as a part of various religious as well as social practices. Cannabis which is a special type of Marijuana can provide temporary relief from analgesia, body pain and in some other clinical conditions. But, impacts of Cannabis on reproductive health of males and females are multi-faceted and differentially fatal. In males, Cannabis can cause changes in testicular morphology, sperm parameters (in terms of semen quality, sperm morphology, sperm mortality and sperm motility), male reproductive hormones and finally causing reduced libido. In females, Cannabis can reduce female fertility by disrupting hypothalamic release of gonadotropin releasing hormone (GnRH), leading to reduced estrogen and progesterone production and anovulatory menstrual cycles. Current research suggest that cannabis may negatively impact on male and female fertility conditions. However, male sterility considering the Cannabis impact is totally lacking in human as well as in sub-human primates. However, very limited studies are available on Cannabis effect on primate female reproduction considering Rhesus monkeys. Hence, further studies are needed to validate that robust findings in animal models will carry over into human experience.

**KEYWORDS:** Cannabis, Effects, Male, Marijuana, Reproduction.

---

Date of Submission: 21-02-2021

Date of acceptance: 04-03-2021

---

### I. INTRODUCTION

*Cannabis* which is a type of marijuana has been used by the people of Indian sub continent from time unknown [1]. They not only use this herb as a part of holy practice but also use it for recreational purposes [2]. Irrespective of sex, this hallucinogenic agent is used by most part of the world particularly by the populations of South America, India, Bangladesh and Pakistan from a long time ago [3]. Reports suggesting the roles of *Cannabis* causing systemic neuropathy [4], neuronal disability [5], impaired fetal development [6] and malfunctioning of male reproductive system [7-10] are documented. But, no reports available are depicting the effects of marijuana in female reproductive system.

The main causative agent of marijuana/cannabinoids is the endocannabinoid. This is a neutral lipid and highly conserved molecule throughout evolutionary history [11]. They are having different derivatives like anandamide [12], 2-arachidonoylglycerol [13] and  $\Delta^9$ -tetrahydrocannabinol (THC) [14]. However, among all of the fatty acid derivatives of cannabinods or endocannabinoids (eCBs) the  $\Delta^9$ -tetrahydrocannabinol (THC) has now been established as the most important hallucinogenic agent of this molecule [15]. There are literatures suggesting the role of this  $\Delta^9$ -tetrahydrocannabinol (THC) in regulation of functions of central nervous system and thus regulating the reproductive functions by affecting/ modulating hypothalamo-pituitary-gonadal axis (HPG-axis) [16] via its receptor CB1 and CB2 [17]. Now it has been reported that CB1 receptors are localized mostly in whole vertebrate central nervous system (CNS) and some peripheral tissues, whereas CB2 receptors are mostly expressed in peripheral tissues and immune cells, however, they have recently been found also in the CNS [18]. But, with all the advancement in psycho-neuro-endocrine research, till date it is a matter of debate how THC is going to regulate reproductive system at peripheral level. Some literatures suggest that, there is a general agreement on the inhibitory effect exerted by cannabinoids and eCBs on GnRH release [19] Thus, it is affecting the subsequent FSH and LH release and impairing both male and female reproduction [20].

In the area of male fertility cannabis use has been linked to reproductive hormone changes, altered semen parameters and reductions in libido and sexual performance [21]. A detailed review of how marijuana affects male fertility at each point along the fertility axis is needed for clinicians to assess the potential risks that patients incur when using this substance.

Although men are more likely to use drugs [22] and have a substance use disorder [23], drug abuse seriously impacts women's health. After alcohol and heroin, marijuana is the most common primary drug of abuse for women entering treatment for substance abuse [24]. Females appear to be more sensitive to the behavioral and physiological effects of marijuana and marijuana-like substances [25], and treatment-seeking

women endorse more severe marijuana withdrawal symptoms than treatment-seeking men [26]. After tobacco and alcohol, marijuana is the most commonly abused substance by women of childbearing age [22]. According to the 2013 NSDUH, 5.4 percent of pregnant women and 11.4 percent of non-pregnant women ages 15 to 44 are current illicit drug users [22], with marijuana representing 64 to 79 percent of female drug use [22,27-29]. As marijuana becomes more widely legalized, marijuana use by women will likely increase [22].

Thus, with this brief introduction it is evident that any form of marijuana can cause detrimental effect on both male and female reproductive health. We have gone through several literatures from Pubmed/Pubmed Central (PMC) and MedLine and with the results procured from our previous experiments on impact of Cannabis on female reproduction [as published elsewhere; 30] the present article is designed. However, the present article is restricted to elucidate the roles of cannabis on male reproduction since the role of cannabis on female reproduction is a matter for another full length chapter. This article may through some light and put forward some recent knowledge on impacts of marijuana and Cannabis in particular on reproductive impairment in males.

## **II. CANNABIS EFFECT ON MALE REPRODUCTION:**

Impacts of Cannabis consumption on male reproduction is multi-dimensional and can be exerted at many levels. Thus, it can be discussed under different sub headings.

### **2. 1. Cannabis effect on sperm parameters:**

#### **2.1.1. Sperm count and concentration:**

Cannabis use is strongly associated with reductions in sperm count and concentration in animal and human studies. Decreased epididymal sperm concentrations were observed in mature male rats exposed to 16 puffs per day of marijuana, comparable to the recreational level in humans, for 75 days [31]. This effect was replicated in a study in which 3 to 6 mg/kg of the Cannabis sativa derivative bhang was administered in adult male mice, which demonstrated a significantly depressed sperm count [32]. Human studies have shown similar findings. A Danish cohort study on marijuana use in 1,215 participants revealed similar changes [33]. Men who reported using marijuana more than once per week had a 28% lower sperm concentration and a 29% lower sperm count than men who had never used marijuana. In a study in which 16 chronic marijuana smokers were exposed to 4 weeks of high dose marijuana the time to a reduced sperm count was 5 to 6 weeks after initiating marijuana use [34]. In human and animal models cannabis has shown strong links to reduced sperm count and concentration which may be linked to arrested spermatogenesis. Future work is needed to elucidate causal mechanisms.

#### **2.1.2. Morphology:**

Cannabis use also appears to induce considerable morphological changes in sperm. In a 1978 study Zimmerman et al treated male mice for 5 consecutive days with intra-peritoneal injections of the marijuana components THC, cannabinoid or cannabitol [35]. On microscopy 35 days after exposure mice treated with THC and cannabitol had a significantly higher incidence of abnormal morphology than the control group, such as banana-shaped, amorphous, folded or hookless heads. Despite morphological changes the research suggests that cannabis does not induce chromosome breakage in sperm. Generoso et al administered 50 mg/kg THC 5 times per week for 6 weeks in 498 male mice [36]. After mating them with females no increase was observed in fetal dominant lethal mutations or heritable translocations over those in controls. These findings were supported by a study by Berryman et al, who found no THC induced increase in pre implantation loss, fetal mortality or the mutation index in fetuses fathered by male mice chronically dosed with THC [37]. In animal and human models evidence suggests that cannabis induces morphological changes in sperm while genetic material is preserved.

#### **2.1.3. Motility and Energy Metabolism:**

The most extensive body of evidence for cannabis related alterations to sperm is for sperm motility. Whan et al noted spermatotoxic effects of THC by incubating sperm with THC at therapeutic doses, similar to concentrations shown to relieve pain or reduce spasticity in humans with multiple sclerosis (0.032 mM) and recreational concentrations (4.8 and 0.32 mM) for 3 hours, and then measuring motility by computer assisted semen analysis [38]. Therapeutic and recreational THC concentrations also resulted in reduced straight line velocity. A similar decrease in sperm motility was seen in an analysis of semen samples from 16 healthy, chronic marijuana users after 4 weeks of high dose marijuana [34]. Barbonetti et al elucidated the mechanism of these findings by establishing a link between CB1 and sperm mitochondrial activity [39]. In sperm incubated with the CB1 receptor agonist Met-AEA a significant reduction was observed in mitochondrial trans-membrane potential. When the sperm were placed under glycolysis blockade, causing them to switch oxidative phosphorylation, the introduction of Met-AEA abolished sperm motility. The link between cannabis and sperm

mitochondrial dysfunction was furthered by Badawy et al [40], who added THC to semen and measured the oxygen concentration as a marker of respiration. Upon the addition of THC mitochondrial respiration immediately declined and was concentration dependent in effect. The results were much more pronounced in washed sperm than in neat semen, suggesting that seminal plasma contains some protective factors. These various investigations suggest that through the action of cannabis on the CB1 receptor the mitochondrial activity of sperm is reduced and as a result motility is significantly impaired. Although Met-AEA and THC administration in the laboratory has helped map potential pathways, to our knowledge it is not known whether these effects are fully replicated in the male testes. Future testing should be done to explore whether mitochondrial impairment is present in the semen of chronic cannabis users.

#### **2.1.4. Viability:**

Cannabis also has a detrimental effect on sperm viability. Rossato et al incubated semen samples with the endocannabinoid AEA at varying concentrations and found that viability was decreased in a dose dependent manner at supraphysiological AEA concentrations [41]. Reduced sperm viability related to cannabis has also been investigated using the highly specific CB1 receptor antagonist rimonabant (SR141716). Cobellis et al found that adding a micromolar concentration of rimonabant induced a small but significant increase in the number of viable spermatozoa [42]. Aquila et al reported similar findings with 1 and 10 nM concentrations of rimonabant increasing sperm viability with no further viability changes observed at higher concentrations [43]. While the cannabinoid system has clear links to sperm viability, future work should be done to confirm these findings with exogenous cannabinoids as well as in the *in vivo* setting.

#### **2.1.5. Fertilization Capacity:**

Research suggests that the cannabinoid signaling pathway may be involved in inhibiting sperm capacitation and activation. Using high performance liquid chromatography Schuel et al observed that high levels of AEA are present in seminal plasma and in progressively decreasing amounts in oviductal and follicular fluid, indicating that sperm are exposed to progressively decreasing AEA levels along the entire fertilization path [44, 45]. The authors speculated that high AEA levels maintain sperm in a quiescent state and the decrease in AEA levels which occurs in the fertilization environment enables sperm to become activated. These data suggest that increases in cannabinoid levels may interfere with sperm activation and may be especially pertinent in the female reproductive tract, which the sperm depend on for tightly regulated AEA levels to maintain proper function. Rossato et al reported that AEA inhibits the capacitation induced acrosome reaction of human sperm after incubation in capacitating medium [41]. Using boar sperm Maccarrone et al found that Met- AEA reduced sperm capacitation in a time dependent manner [46]. They also noted that this effect was mediated by the CB1 receptor since when rimonabant, which blocks CB1, was added, Met-AEA produced no change in capacitation. Schuel et al used the cannabinoid agonist AM-365 to identify concentration dependent stimulation and inhibition of sperm hyperactivated motility, which is a state needed for sperm to reach the egg surface and which assists with penetration of the zona pellucida. Current work suggests that the endocannabinoid system is intimately linked to the fertilization process in the male and female reproductive tracts. Given the well described inhibitory effects, cannabis is likely to have negative impacts on fertilizing potential.

### **III. CANNABIS EFFECTS ON REPRODUCTIVE HORMONES:**

#### **3.1. Follicle Stimulating Hormone:**

Relatively few studies have focused on cannabis and FSH levels, and most have observed no effect. Cone et al found no significant change in FSH levels in 4 healthy males with a history of frequent marijuana use before and after 2 marijuana cigarettes per day for 3 consecutive days [47]. Vescovi et al observed that cannabis did not alter the response of FSH to gonadotropin-releasing hormone in 10 male chronic marijuana users given gonadotropin-releasing hormone intravenously [48]. A depression in FSH levels was observed only by Kolodny et al, who compared plasma hormone levels among 11 men who used 5 to 9 marijuana cigarettes per week, 9 who used 10 or more marijuana cigarettes per week and normal controls [29]. Based on current studies, FSH may not be affected by cannabis except perhaps in the limited case of heavy chronic use. Human studies to date have been limited in suggestive power due to the small cohort sizes, leaving considerable room for further validation with larger sample size investigations.

#### **3.2. Luteinizing Hormone:**

In human and animal models LH is consistently lowered by cannabis [49, 50]. In the single exception Kolodny et al did not observe any significant difference in plasma LH levels between men who smoked 5 to 9 marijuana cigarettes per week and men who smoked 10 or more per week.12 However, the variation in marijuana use levels in this study may have been insufficient to induce LH variations. The relationship between

cannabis and LH was strengthened in a study by Wenger et al, who used polyclonal antibodies against CB1 and CB2 to localize individual cells expressing cannabinoid receptors [51]. The CB1 receptor was found in the anterior pituitary in LH secreting gonadotrophic cells. Wenger et al reaffirmed these results after administering AEA to wild-type and CB1 knockout mice, which revealed decreased LH secretion in the wildtype mice but unchanged LH levels in the CB1 knockout mice.<sup>32</sup> As is the case with FSH related investigations, understanding how cannabis impacts LH would be improved by larger randomized, controlled trials in human subjects.

### **3.3. Testosterone:**

The reported effect of cannabis on serum testosterone levels is widely variable across current studies. In an early work in 20 chronic marijuana users Kolodny et al found a significant reduction in testosterone levels between chronic and never marijuana users [29]. The evidence that cannabis depresses testosterone levels relies heavily on animal studies. In contrast to findings in animals, most human studies support the conclusion that testosterone levels are not significantly changed by cannabis use. A 1974 study by Mendelson et al in 27 chronic marijuana users who were administered marijuana for 21 days showed no significant changes in plasma testosterone levels.<sup>34</sup> In a 1986 study of 4 male chronic marijuana users given 2 marijuana cigarettes per day depressed free testosterone levels were observed but the levels did not significantly differ from baseline [47]. In a later study free testosterone levels were compared in 41 normal controls and in 66 Pakistani men who smoked cannabis daily or regularly consumed cannabis tea.<sup>35</sup> No significant difference was observed in plasma testosterone levels between the cannabis users and the normal controls. Although sample size was limited in these early human studies, they suggest that cannabis consumption does not significantly alter testosterone levels. It is only recently that large cohort studies of cannabis users have been possible. To date these studies have continued the trend of presenting conflicting or inconclusive evidence on the link between cannabis use and testosterone levels.

The first large cohort study on the effects of cannabis use was performed by Gundersen et al in 2015 using a registry of 1,215 Danish men undergoing compulsory medical examination to determine fitness for service [33]. Testosterone levels were 7% higher in self-reported marijuana smokers than in nonusers. This was within the same range of testosterone elevation observed in cigarette smokers in the cohort. The authors cautioned that the increased testosterone levels in marijuana users could not be separated from the effect of tobacco smoking alone. A second major cohort study was done in 2017 by Thistle et al [53]. They used data on 1,577 American men using data from the 2011 to 2012 United States National Health and Nutrition Survey with several novel outcomes. No difference was observed in serum testosterone levels between ever and never users of marijuana. However, serum testosterone levels showed an inverse association with time since the last regular use of marijuana, and since the last marijuana use.

This indicated that recency rather than frequency of use may have the strongest relationship with serum testosterone levels. Additional large, population based samples are needed to clarify currently conflicting reported effects of cannabis on testosterone levels.

## **IV. CONCLUSION:**

As cannabis increasingly gains legalized status across the United States, the popularity and prevalence of use continue to grow. Although medically it demonstrates therapeutic promise in some areas such as multiple sclerosis and chronic neuropathic pain, the potential adverse effects remain widely under studied. Current research shows that cannabis likely has negative impacts at several points along the male fertility pathway. Human sperm express cannabinoid receptors, suggesting that they are directly impacted by alterations in the balance of the endocannabinoid system. The effect of cannabis on testosterone levels is largely undetermined while LH levels appear to be lowered and FSH levels are unchanged. The strongest evidence for the deleterious effects of cannabis on male reproductive capacity is its impact on semen parameters. Studies demonstrate reduced sperm count and concentration, morphological

changes, reduced motility and viability, and decreased fertilizing capacity in animals and humans exposed to marijuana or cannabis derivatives. Furthermore, animal studies suggest that cannabis has a role in testicular atrophy. While cannabis may increase libido in the short term, chronic use may diminish erectile function in men. The evidence presented to date largely relies on animal models, *in vitro* studies of endogenous cannabinoid compounds and retrospective analyses. The ethical and legal complications of an *in vivo*, controlled study drive the limited amount of data presented in human subjects, a limitation which is unlikely to abate going forward. Future studies should focus on gathering large cohort data in national surveys, similar to the voluntary, cannabis related data collection done with compulsory military fitness examinations in different countries. These types of studies are needed to confirm that animal models can be translated to human experience.

REFERENCES:

- [1]. Ashton CH, Moore PB, Gallagher P, Young AH. Cannabinoids in bipolar affective disorder: a review and discussion of their therapeutic potential. *Psychopharmacol.* 2005; 9:293.
- [2]. Aversa A, Rossi F, Francomano D, et al. Early endothelial dysfunction as a marker of vasculogenic erectile dysfunction in young habitual cannabis users. *Int J Impot Res.* 2008; 20:566.
- [3]. Battista N, Pasquariello N, Di Tommaso, Maccarrone M. Interplay between endocannabinoids, steroids and cytokines in the control of human reproduction. *J Neuroendocrinol.* 2008; 20:82.
- [4]. Bayewitch M, Rhee MH, Avidor-Reiss T, et al. D<sup>9</sup>-Tetrahydrocannabinol antagonizes the peripheral cannabinoid receptor-mediated inhibition of adenylyl cyclase. *J Biol Chem.* 1996; 1271:9902.
- [5]. Brown TT, Dobs AS. Endocrine effects of marijuana. *J Clin Pharmacol.* 2002; 42:S90.
- [6]. Cacciola G, Chioccarelli T, Mackie K, et al. Expression of type-1 cannabinoid receptor during rat postnatal testicular development: possible involvement in adult Leydig cell differentiation. *Biol Reprod.* 2008; 79:758.
- [7]. Cobellis G, Cacciola G, Scarpa D, et al. Endocannabinoid system in frog and rodent testis: type-1 cannabinoid receptor and fatty acid amide hydrolase activity in male germ cells. *Biol Reprod.* 2006; 75: 82.
- [8]. Chakravarty I, Shah PG, Sheth AR, Ghosh JJ. Mode of action of delta-9-tetrahydrocannabinol on hypothalamo-pituitary function in adult female rats. *J Reprod Fertil.* 1979; 57:113.
- [9]. Cravatt BF, Giang DK, Mayfield SP, et al. Molecular characterization of an enzyme that degrades neuromodulatory fatty-acid amides. *Nature.* 1996; 384:83.
- [10]. Campbell VA. Tetrahydrocannabinol-induced apoptosis of cultured cortical neurones is associated with cytochrome c release and caspase-3 activation. *Neuropharmacol.* 2001; 40:702.
- [11]. Dalterio S, Bartke A, Burstein S. Cannabinoids inhibit testosterone secretion by mouse testes *in vitro*. *Science.* 1977; 196:1472.
- [12]. Singh SK, Chakravarty S. Antispermatic and antifertility effects of 20, 25-diazacholesterol dihydrochloride in mice. *Reprod Toxicol.* 2003; 17:37.
- [13]. Devane WA, Hanus L, Breuer A. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science.* 1992; 258:1946.
- [14]. Dhawan K, Kumar S, Sharma A. Reversal of cannabinoids (delta9- THC) by the benzoflavone moiety from methanol extract of *Passiflora incarnata* Linnaeus in mice: a possible therapy for cannabinoid addiction. *J Pharm Pharmacol.* 2002; 54:875.
- [15]. Dixit VP, Lohiya NK. Effects of Cannabis extract on the response of accessory sex organs of adult male mice to testosterone. *Ind J Physiol Pharmacol.* 1975; 19:98.
- [16]. Dixit VP, Gupta CL, Agarwal M. Testicular degeneration and necrosis induced by chronic administration of cannabis extract in dogs. *Endokrinologie.* 1977; 69:299.
- [17]. EL-Gohary M, Eid MA. Effect of cannabinoid ingestion (in the form of bhang) on the immune system of high school and university students. *Hum Exp Toxicol.* 2004; 23:149.
- [18]. Gammon CM, Freeman Jr GM, Xie W, et al. Regulation of gonadotropin-releasing hormone secretion by cannabinoids. *Endocrinol.* 2005; 146:4491.
- [19]. Gaoni Y, Mechoulam R. Isolation, structure and partial synthesis of an active constituent of hashish. *J Am Chem Soc.* 1964; 86:1646.
- [20]. Gye MC, Kang HH, Kang HJ. Expression of cannabinoid receptor 1 in mouse testes. *Arch Androl.* 2005; 51:247.
- [21]. Bari M, Battista N, Pirazzi V et al: The manifold actions of endocannabinoids on female and male reproductive events. *Front Biosci (Landmark Ed)* 2011; 16: 498.
- [22]. Substance Abuse and Mental Health Services Administration. Results from the 2013 National Survey on Drug Use and Health: Summary of National Findings. Rockville, MD: Substance Abuse and Mental Health Services Administration; 2014. Report No.: NSDUH Series H-48, HHS, Publication No. (SMA) 14-4863 [internet]. Available from: <http://www.samhsa.gov/data/sites/default/files/NSDUHresultsPDFwHTML2013/web/NSDUHresults2013.pdf>
- [23]. Grant BF, Saha TD, Ruan wJ, Goldstein RB, Chou SP, Jung J, Zhang H, Smith SM, Pickering RP, Huang B, Hasin DS. Epidemiology of DSM-5 drug use disorder: Results from the national epidemiologic survey on alcohol and related conditions- iii. *JAMA Psychiatry.* 2015;18:1-9.
- [24]. Substance Abuse and Mental Health Services Administration, Center for Behavioral Health Statistics and Quality. Treatment Episode Data Set (TEDS): 2002-2012. National Admission to Substance Abuse Treatment Services. Rockville, Maryland: SAMHSA; 2014. Report No.: BHSIS Series S- 71, HHS Publication No. (SMA) 14-4850.
- [25]. Craft RM. Sex differences in behavioral effects of cannabinoids. *Life Sci.* 2005;77(20):2471.
- [26]. Herrmann ES, weerts EM, vandrey R. Sex differences in cannabis withdrawal symptoms among treatment-seeking cannabis users. *Exp Clin Psychopharmacol.* 2015;23:415.
- [27]. Sherwood RA, Keating J, Kavvadia v, Greenough A, Peters TJ. Substance misuse in early pregnancy and relationship to fetal outcome. *Eur J Pediatr.* 1999;158:488.
- [28]. Garcia-Serra J, Ramis J, Simo S, Joya X, Pichini S, vall O, Garcia-Algar O. Alternative biological materials to detect prenatal exposure to drugs of abuse in the third trimester of pregnancy. *An Pediatr (Barc).* 2012;77:323.
- [29]. Ebrahim SH, Gfroerer J. Pregnancy-related substance use in the united states during 1996-1998. *Obstet Gynecol.* 2003;101:374.
- [30]. Ghosh S, Rai SK. Chronic cannabis induced oxidative stress and reproductive containment in female mice. *Int J Green Pharm.* 2018. 12.
- [31]. Huang HF, Nahas GG and Hembree WC 3rd: Effects of marihuana inhalation on spermatogenesis of the rat. *Adv Biosci* 1978; 22-23: 419.
- [32]. Banerjee A, Singh A, Srivastava P et al: Effects of chronic bhang (cannabis) administration on the reproductive system of male mice. *Birth Defects Res B Dev Reprod Toxicol* 2011; 92: 195.
- [33]. Gundersen TD, Jørgensen N, Andersson AM et al: Association between use of marijuana and male reproductive hormones and semen quality: a study among 1,215 healthy young men. *Am J Epidemiol* 2015; 182: 473.
- [34]. Hembree WC 3rd, Nahas GG, Zeidenberg P et al: Changes in human spermatozoa associated with high dose marihuana smoking. *Adv Biosci* 1978; 22-23: 429.
- [35]. Zimmerman AM, Zimmerman S and Raj AY: Effects of cannabinoids on spermatogenesis in mice. *Adv Biosci* 1978; 22-23: 407.
- [36]. 17. Generoso WM, Cain KT, Cornett CV et al: Tests for induction of dominant-lethal mutations and heritable translocations with tetrahydrocannabinol in male mice. *Mutat Res* 1985; 143: 51.
- [37]. Berryman SH, Anderson RA Jr, Weis J et al: Evaluation of the co-mutagenicity of ethanol and delta 9-tetrahydrocannabinol with Trenimon. *Mutat Res* 1992; 278: 47.

- [38]. Whan LB, West MC, McClure N et al: Effects of delta-9-tetrahydrocannabinol, the primary psychoactive cannabinoid in marijuana, on human sperm function in vitro. *Fertil Steril* 2006; 85: 653.
- [39]. Barbonetti A, Vassallo MR, Fortunato D et al: Energetic metabolism and human sperm motility: impact of CB1 receptor activation. *Endocrinology* 2010; 151: 5882.
- [40]. Badawy ZS, Chohan KR, Whyte DA et al: Cannabinoids inhibit the respiration of human sperm. *Fertil Steril* 2009; 91: 2471.
- [41]. Rossato M, Ion Popa F, Ferigo M et al: Human sperm express cannabinoid receptor Cb1, the activation of which inhibits motility, acrosome reaction, and mitochondrial function. *J Clin Endocrinol Metab* 2005; 90: 984.
- [42]. Cobellis G, Cacciola G, Scarpa D et al: Endocannabinoid system in frog and rodent testis: type-1 cannabinoid receptor and fatty acid amide hydrolase activity in male germ cells. *Biol Reprod* 2006; 75: 82.
- [43]. Aquila S, Guido C, Santoro A et al: Human sperm anatomy: ultrastructural localization of the cannabinoid1 receptor and a potential role of anandamide in sperm survival and acrosome reaction. *Anat Rec (Hoboken)* 2010; 293: 298.
- [44]. Schuel H and Burkman LJ: A tale of two cells: endocannabinoid-signaling regulates functions of neurons and sperm. *Biol Reprod* 2005; 73: 1078.
- [45]. Schuel H, Burkman LJ, Lippes J et al: Evidence that anandamide-signaling regulates human sperm functions required for fertilization. *Mol Reprod Dev* 2002; 63: 376.
- [46]. Maccarrone M, Barboni B, Paradisi A et al: Characterization of the endocannabinoid system in boar spermatozoa and implications for sperm capacitation and acrosome reaction. *J Cell Sci* 2005; 118: 4393.
- [47]. Cone EJ, Johnson RE, Moore JD et al: Acute effects of smoking marijuana on hormones, subjective effects and performance in male human subjects. *Pharmacol Biochem Behav* 1986; 24: 1749.
- [48]. Wenger T, Rettori V, Snyder GD et al: Effects of delta-9-tetrahydrocannabinol on the hypothalamic-pituitary control of luteinizing hormone and follicle-stimulating hormone secretion in adult male rats. *Neuroendocrinology* 1987; 46: 488.
- [49]. Vescovi PP, Pedrazzoni M, Michelini M et al: Chronic effects of marijuana smoking on luteinizing hormone, follicle-stimulating hormone and prolactin levels in human males. *Drug Alcohol Depend* 1992; 30: 59.
- [50]. Martini-Calderon JL, Muñoz RM, Villanueva MA et al: Characterization of the acute endocrine actions of (-)-11-hydroxy-delta8-tetrahydrocannabinol-dimethylheptyl (HU-210), a potent synthetic cannabinoid in rats. *Eur J Pharmacol* 1998; 344: 77.
- [51]. Wenger T, Fernandez-Ruiz JJ and Ramos JA: Immunocytochemical demonstration of CB1 cannabinoid receptors in the anterior lobe of the pituitary gland. *J Neuroendocrinol* 1999; 11: 873.
- [52]. Mendelson JH, Kuehnle J, Ellingboe J et al: Plasma testosterone levels before, during and after chronic marijuana smoking. *N Engl J Med* 1974; 291: 1051.
- [53]. Thistle JE, Graubard BI, Braunlin M et al: Marijuana use and serum testosterone concentrations among U.S. males. *Andrology* 2017; 5: 732.

Somenath Ghosh\*. "Male reproduction and Cannabis: A state of art review." *International Journal of Pharmaceutical Science Invention*, vol. 10(03), 2021, pp 04-09. Journal DOI- 10.35629/6718